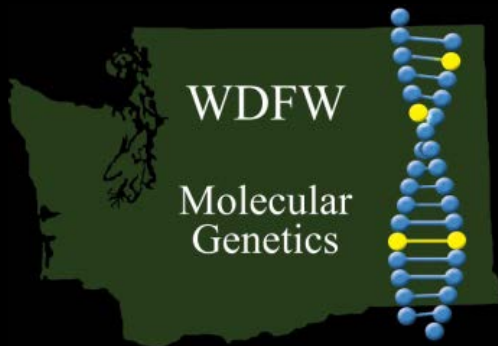


DNA-Based Pedigree Assignments of Chinook Salmon from the Yakima River

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Background

- Joint project between WA Department of Fish and Wildlife (WDFW) and Yakama Nation (YN)
- Project objective is to assess the relative fitness of natural-origin Chinook with different percentages of hatchery ancestry in natural environment of the upper Yakima River
- Collection of hatchery-origin adult males and females, jacks, and precocious male Chinook at Roza Dam 2003 – 2006; hatchery- and natural-origin Chinook 2007 - present
- Genetic analysis using microsatellite DNA loci follows the same methodology used for the analysis of Chinook in the Cle Elum spawning channel

Laboratory Methods

- DNA was extracted from fin tissue
- PCR amplification was performed using microsatellite loci
- Amplified products were run through an ABI-3730 Genetic Analyzer
- Electropherograms were scored using GENEMAPPER software v.3.7
- Data was binned using GAPS allele naming

Cherril setting up DNA extraction



Laboratory Methods

- DNA was extracted from fin tissue
- PCR amplification was performed using microsatellite loci
- Amplified products were run through an ABI-3730 Genetic Analyzer
- Electropherograms were scored using GENEMAPPER software v.3.7
- Data was binned using GAPS allele naming

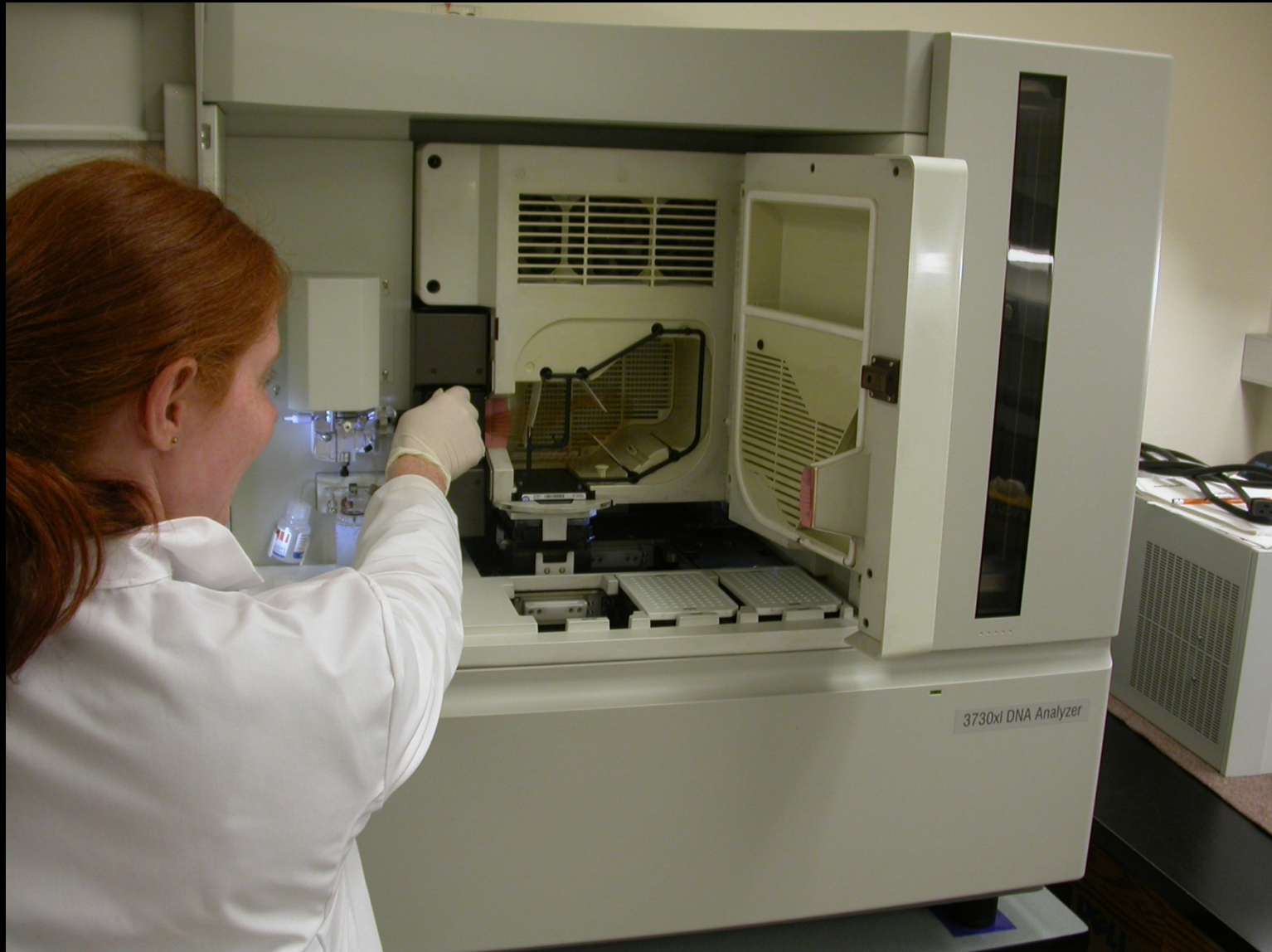
Cheryl setting up PCR reaction



Laboratory Methods

- DNA was extracted from fin tissue
- PCR amplification was performed using microsatellite loci
- Amplified products were run through an ABI-3730 Genetic Analyzer
- Electropherograms were scored using GENEMAPPER software v.3.7
- Data was binned using GAPS allele naming

Jennifer loading the ABI-3730



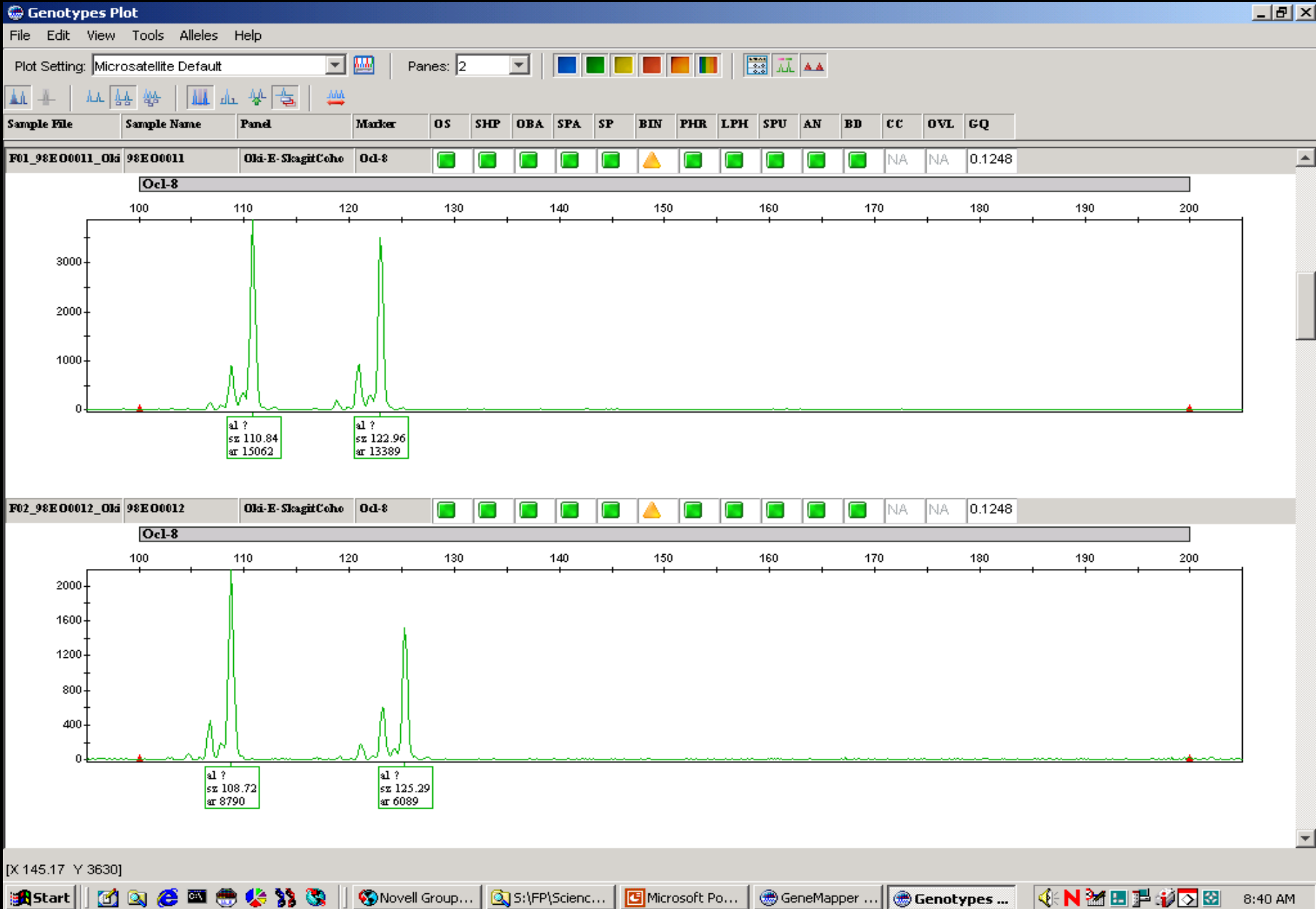
Laboratory Methods

- DNA was extracted from fin tissue
- PCR amplification was performed using microsatellite loci
- Amplified products were run through an ABI-3730 Genetic Analyzer
- Electropherograms were scored using GENEMAPPER software v.3.7 (38,436 individual electropherograms)
- Data was binned using GAPS allele naming

Jennifer Scoring an Electropherogram



Electropherogram – Ocl-8



Laboratory Methods

- DNA was extracted from fin tissue
- PCR amplification was performed using microsatellite loci
- Amplified products were run through an ABI-3730 Genetic Analyzer
- Electropherograms were scored using GENEMAPPER software v.3.0
- Data was binned using GAPS allele naming

Locus Data

Locus	N Alleles	N parents Genotyped	H _o	H _e	Excl (1)	Excl (2)
Ogo-2	11	2,186	0.825	0.821	0.475	0.648
Ogo-4	11	2,188	0.801	0.806	0.456	0.632
Oki-100	26	2,117	0.919	0.904	0.682	0.811
Omm-1080	44	2,162	0.937	0.961	0.852	0.920
Ots-201b	29	2,118	0.915	0.904	0.679	0.809
Ots-208b	29	2,115	0.930	0.941	0.787	0.880
Ots-211	28	2,123	0.930	0.931	0.757	0.861
Ots-212	24	2,182	0.887	0.887	0.631	0.774
Ots-213	29	2,185	0.921	0.936	0.769	0.869
Ots-3M	9	2,185	0.652	0.651	0.254	0.435
Ots-9	6	2,186	0.678	0.656	0.237	0.400
Ots-G474	13	2,190	0.362	0.367	0.072	0.211
Ssa-197	25	2,180	0.902	0.906	0.683	0.812
Ssa-408	27	2,160	0.728	0.916	0.709	0.830

Excl (1) = Exclusionary ability of the locus when neither parent is known

Excl (2) = Exclusionary ability of the locus when one parent is known

Evaluation of Parentage Assignments

- Maximum likelihood parentage assignments performed with the program CERVUS 3.0
- Assignments for offspring were calculated for the two most likely male and female parent pair. The parent pair assignment with two mismatches or less was accepted
- Individuals that did not assign to a parent pair were then analyzed for a female parent only and male parent only (assignments with two or less mismatches were accepted)
- Maximum likelihood analysis was then conducted using the program MYKISS
- An additional 22 individuals were assigned to a parent pair

Causes of Mismatching

- Germ-line mutation – a parent passes a changed allele to their offspring (sequence or allele changes during replication)
- PCR error (or process error) – error introduced by poor amplification from lower quality DNA extracts
- Genotyping error – inadvertent human error and computer software error in scoring due to multiple peaks being selected

Electropherogram – Oki-100



Mismatching

	Oki-100	Ots-3M	Ots-213
Female – 1	100/100	100/100	100/100
Female – 2	200/200	200/200	200/200
Male – 1	120/120	120/120	120/120
Male – 2	240/240	240/240	240/240
Offspring – 1	100/120	100/120	100/120
Offspring – 2	200/240	200/240	200/240
Offspring – 3	100/120	100/120	100/240

Expected proportion - Hatchery- and Natural-origin Chinook

- 2,284 – Hatchery-origin Chinook count at Roza Dam
 - 1,558 / 1,147 – Natural-origin Chinook count at Roza Dam
(411 – Natural-origin Chinook brood)
 - $2,284/3,431 = 0.6657 - P$; $1,147/3,431 = 0.3343 - Q$
-
- 44.3% Hatchery-origin (H X H) – P^2
 - 44.5% Hatchery & Natural-origin (H X N & N X H) – $2PQ$
 - 11.2% Natural-origin (N X N) - Q^2

Observed returns - Hatchery- and Natural-origin Chinook

- **161 / 569** offspring were assigned parental pair
Hatchery X Hatchery (28.3%)
- **230 / 569** offspring were assigned a mother only
Hatchery X Natural (40.4%)
- **62 / 569** offspring were assigned a father only
Natural X Hatchery (10.9%)
- **116 / 569** offspring did not assign a male or female
Natural X Natural (20.4%)

Comparison of Expected and Observed Percentages of Hatchery and Natural-Origin Chinook

	Expected	Observed
H X H	45.0%	28.3%
H X N & N X H	44.0%	51.3%
N X N	11.0%	20.4%

Conclusions

- Preliminary data
- The number of observed natural-origin Chinook exceeds the expected amount while hatchery-origin is lower than expected
- More female only assignments than male assignments of hatchery-origin

Future Work

- Analysis of 1999 brood to determine an error rate for calculating N X N offspring in the 2007 collection
- Analysis of remaining 2007 four year Chinook
- Analysis of 2004 adults and 2008 offspring
- Analysis of 2011 and 2012 returns

Acknowledgements

- BPA funds for the YKFP supported this work effort
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